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The impact of growth promoters on muscle growth and the potential consequences for meat quality.

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Abstract

To meet the demands of increased global meat consumption, animal production systems will have to become more efficient, or at least maintain the current efficiency utilizing feed ingredients that are not also used for human consumption. Use of growth promoters is a potential option for increasing production animal feed efficiency and increased muscle growth. The objective of this manuscript is to describe the mechanisms by which the growth promoters, beta-adrenergic agonists and growth hormone, mediate their effects, with specific consideration of the aspects which have implications for meat quality.

1. Introduction

The predicted increase in world population to 9 billion by 2050 is expected to be associated with an increase in the urban population, with an estimated 70% being in an urban environment compared to 49% today and, hopefully, the inevitable increase in incomes (FAO, 2009). To meet this demand it is estimated that the food production will have to increase, particularly for commodities associated with higher incomes, such as meat, with the predicted demand more than doubling by 2050 (FAO, 2009).

To date, significant advances in animal genetics and animal nutrition have been made to meet the increasing demand. To achieve the maximum growth potential, high quality feed ingredients are required. Feed ingredients account for a large proportion of the overall costs of animal production, particularly in non-ruminant species (Patience, Rossoni-Serao, & Gutierrez, 2015). Continuing to rely on the same ingredients, in competition with human nutrition and biofuels, mean prices will inevitably increase. Therefore the cost of meat and animal products will also increase. It has been estimated that for many agricultural commodities the rate of production has already reached a peak (Seppelt, Manceur, Liu, Fenichel, & Klotz, 2014). Hence, if we are to continue to meet the demand for animal products, we cannot simply feed more animals the same feed ingredients, as that would require more crops, land and water (Foresight, 2011; Godfray et al., 2010). Therefore the aim of current research is to improve the efficiency with which animals utilise their feeds, giving more product for the same amount of feed or the same amount of product for less feed.

Through selective breeding and improved diet formulations over the last 20-30 years, feed efficiency of pigs (Patience et al., 2015) and chickens (Siegel, 2014) has improved, with Feed Conversion Ratio (FCR) values of 2.0 or less currently achievable (i.e. >50% efficiency). The UN suggests productivity is likely to be enhanced in the future through better animal disease control, improved irrigation and water management practices, and better fertilizers

(FAO, 2009). In terms of animal production the increase in productivity could also be increased through the continued utilization of genetic selection through breeding programmes. Also it is probably inevitable that molecular biology technologies will be accepted and GMO organisms will be utilized for animal production. However throughout the world there is an increasing utilization of growth promoters. A goal of all these technologies is to increase the efficiency of feed utilization that ideally results in an increase in lean carcass weight. A predominant group of growth promoters are those which are based on endocrine factors such as anabolic steroids, growth hormone (GH, also called somatotropin, ST) and beta-adrenergic agonists (BA). These agents have metabolic modifying characteristics that result in enhanced growth. In addition there are a variety of other types of growth promoters, such as antibiotics, whose predominant affect is thought to be the increased efficiency of utilization of nutrients from the gut. Although the use of growth promoting agents is banned in the European Union (EU), they are legally used in many other countries.

Increases in lean tissue deposition (increased muscle mass), decreases in fat deposition, along with repartitioning nutrients away from fat to muscle growth, are some of the predominant objectives of animal production systems focused on generating meat. Therefore these are key features of animal growth that growth promoters are designed to influence. The actions of hormonal based growth promoters' are through mechanisms that influence processes at a cellular level. In addition to affecting protein turnover to increase protein accretion, they also have effects on energy metabolism. This manuscript will examine the effects of hormonal based growth promoters on carcass parameters that influence meat quality, particularly growth hormone (GH) and beta-adrenergic agonists (BA), and will predominantly focus on their effects in pigs.

2. Growth Hormone

Growth hormone is a peptide hormone produced by the anterior pituitary and is involved in the processes of development and growth that includes skeletal muscle, bone and adipose tissue (Beermann, 1994). Increased plasma GH has the effect of redirecting nutrients away from adipose tissue and toward muscle and bone (Etherton & Bauman, 1998). The effect of GH is to stimulate the production of insulin like growth factor (IGF-1) from the liver. These observations led to the hypothesis that GH effects, particularly on growth, were mediated by IGF-1 originating from the liver (Daughaday, 2000). However GH can stimulate IGF-1 expression in other tissues, particularly bone but also skeletal muscle (Brameld et al., 1996; Velloso, 2008: Wang, Bikle & Chang, 2013). In the circulation IGF-1 is associated with

insulin like growth factor binding proteins (IGFBP) which prolong its half-life (Boisclair, Rhoads, Ueki, Wang, & Ooi, 2001). The effect of IGF-1 can be mediated through dimers of the IGF-1 receptor as well as IGF-1 receptor/insulin receptor hybrids (Denley, Cosgrove, Booker, Wallace, & Forbes, 2005) and subsequently activates multiple signalling pathways. In muscle the predominant pathways are the mitogen-activated protein kinase kinase/extracellular signal-regulated kinase (MEK/ERK pathway, which tends to be associated with proliferative growth, whilst the protein kinase B -mechanistic target of rapamycin - ribosomal protein S6 kinase (Akt-mTOR-S6K) pathway is predominantly responsible for protein synthesis but also affects protein degradation (Clemmons, 2009). Growth Hormone is approved for use in 14 countries, including Australia, for administration to pigs to improve growth characteristics (Dunshea, Cox, Borg, Sillence, & Harris, 2002). As the GH is a peptide it has to be administered by regular injection. Its action on pig growth has been well characterised, reducing feed intake whilst simultaneously increasing lean and reducing fat deposition (Etherton et al., 1987). Unsurprisingly, due to the signalling pathway activated by GH-IGF-1 axis, increased protein accretion in GH treated pigs is stimulated by protein synthesis rather than a decrease in protein degradation. However there are inconsistent reports in the literature with some reporting similar increases in protein synthesis and breakdown but overall net increased nitrogen balance (Tomas et al., 1992), whilst others have reported that protein synthesis is increased but protein degradation appears not to be affected (Bush et al., 2003) and some have described even a decrease in degradation (Vann et al., 2000). The effect of exogenous GH on adipose tissue is to reduce fat synthesis which results in reductions in back fat (Krick et al., 1992). Dunshea et al. (2005) carried out a comprehensive review on the effects of GH on pig meat quality. Using a meta-analysis approach on published data, they concluded that GH decreased intramuscular fat by 12% and increased shear force by 9%. The effects on shear force are unlikely to be due to a large decrease in the proteolytic activity. Therefore the increased shear force is likely to be due to changes in temperature transfer to the meat, either during chilling the carcass, thereby affecting rigor development or effects of heat transfer during cooking which influences denaturing of proteins. Overall Dunshea et al. (2005) concluded that GH causes a small increase in shear force and the sensory perception of meat from treated pigs is tough, but it was unlikely that that this could be detected by consumers (Dunshea, D'Souza, Pethic, Harper, & Warner, 2005)

3. Beta-adrenergic agonists

The BAs are analogues of the endogenous catecholamines, adrenaline and noradrenaline. When administered to livestock species they have positive effects on growth and nutrient repartitioning. Beta-2 adrenergic receptor specific agonists, such as clenbuterol and cimaterol, have the greatest growth effect; however ractopamine, which binds to both beta-1 and -2 adrenergic receptors (Mills, Kissel, Bidwell, & Smith, 2003), also has similar growth effects. The BA generally have positive effects on weight gain, FCR, and act as strong repartitioning agents, increasing muscle growth whilst decreasing adipose tissue deposition (Meersman, 1998).

The BAs mediate their growth effects through the BA receptor. The subtypes of beta adrenergic receptors (BAR) vary depending on the tissue. For example, in pigs skeletal muscle has more BAR2 than BAR1 whilst in adipose tissue it is the reverse (Liang & Mills, 2002). BAR3 are thought to be present on porcine adipocytes (McNeel & Mersmann, 1995). The BAR activates the adenylate cyclase pathway which subsequently produces cyclic adenosine monophosphate (cAMP), which then activates protein kinase A. This kinase can alter enzyme activity through phosphorylation. For example the phosphorylation dependent cascade results in phosphorylase being activated and glycogen being degraded to glucose-1-phosphate. In addition, protein kinase A can activate the transcription factor, cAMP response element binding protein (CREB), which then regulates the transcription of genes that have a functional cAMP responsive element within their regulatory regions (Altarejos & Montminy, 2011). Therefore these agents have an immediate effect on enzyme activity, but can also alter transcription of a number of genes.

The first BA to be licensed for use as a feed additive was ractopamine for use in pigs in 1999. A significant advantage of these agents when compared to GH is that they can be administered in feed. The predominant effect of BA is to increase lean deposition whilst also decreasing fat deposition (Meersman, 1998). The BAs have strong effects on muscle hypertrophy and appear to be differentially effective across farm species, with ruminant species (cattle and sheep) responding the strongest, being particularly effective in older animals (Meersman, 1998). The early studies on the effects of BA in livestock indicated that these agents had a strong effect of decreasing protein degradation without increasing protein synthesis (Bohorov, Buttery, Correia, & Soar, 1987). However subsequent studies in pigs treated with ractopamine have indicated that protein synthesis, particularly of myofibrillar proteins, is stimulated (Adeola, Ball, & Young, 1992). The effects on fat deposition are not as clear as for GH, particularly in pigs, however it appears that ractopamine reduces backfat thickness in pigs, but this is not as dramatic as the effect of BA in ruminant species (Dunshea

et al., 2005). After reviewing a large number of studies, Dunshea et al. (2005) concluded that the use of BA in ruminants increased the shear force, with cimaterol increasing shear force by 60%. Using a meta-analysis approach on published data they also described how, in pigs, the BAs ractopamine and salbuterol had no effect on IMF, whilst cimaterol caused a decrease. All these BA had the effect of increasing shear force in pigs, but again cimaterol had the greatest negative effect, but the effects were not as great as in ruminants. Unlike GH, the effect of BA on tenderness appears to be mediated through their strong inhibitory effects on protein degradation (see below), rather than effects on IMF.

4. Fibre type and meat quality

Skeletal muscle is made up of muscle fibres which have differing contractile and metabolic characteristics. Muscle fibres can be histochemically classified according to their actomyosin ATPase activity into three types, types I, IIA and IB, which have a contractile and associated metabolic characteristics of slow twitch oxidative, fast twitch oxidative and fast twitch glycolytic, respectively (Brooke & Kaiser, 1970). When considered at a molecular level the differences in fibre type are, in part, determined by the myosin heavy chains (MyHC) they express, as this protein is a determinant of muscle contractile speed and thereby the associated metabolism required to support this. There are four MyHC isoforms (types I, IIa, IIx and IIb) which has led to a revision in the classification of fibre types as types I, IIA, IIX and IIB with a rank order of contraction speed of I<IIA<IIX<IIB (Schiaffino & Reggiani, 1996). The MyHCIIb isoform is expressed in rodents, but absent from most large animals such as cattle and sheep, although pigs do express this isoform (Lefaucheur, 2009). To support the contraction of these fibres there are associated metabolic characteristics. At one extreme, the slow twitch fibres that express high levels of MyHCI protein have a dependence on oxidative (aerobic) metabolism, as they can maintain a low power output. These fibres have a good blood supply and tend to have a low glycogen and high triglyceride content and fatigue slowly, so are used for sustained contraction. In contrast, the fast twitch type II fibres have a dependence on glycolytic (anaerobic) metabolism. These fibres have a relatively low blood supply and have a high glycogen content and low triglyceride content and fatigue relatively quickly so are used for rapid, intense contraction, which cannot be sustained for long periods. The type IIB fibres have the highest glycolytic metabolism capacity. The type IIA and IIX fibres have metabolic characteristics that, although still enable a high degree of glycolytic metabolism, increasingly have the capacity to carry out oxidative metabolism, hence they are classified as oxidative-glycolytic, particularly the type IIA fibres which have the greatest

aerobic capacity (Schiaffino & Reggiani, 1996). For glycolytic fibres (type IIA, IIX and IIB) there is not a simple relationship between the expression of the fast MyHC protein isoforms (IIa, IIx and IIb) and the fibre type classification based on contractile and metabolic characteristics. For example in Large White pig longissimus muscle, fibres classified as type IIB included MyHC IIx expressing fibres, as well as hybrid fibres expressing both MyHC IIx and IIb (Lefuacher, 2009). These hybrid fibres are a particular characteristic of the fast glycolytic fibres, which means using MyHC expression data to determine fibre type (as normally defined by contractile and metabolic characteristics), should be done with caution.

Based on the association between the metabolic characteristics of muscle and fibre type, it would be expected that IMF content would be lower in muscles that contain a high proportion of fast fibre types, particularly those containing a high proportion of type IIB fibres, for example in pigs. Recent work in pigs has indicated no clear relationship between proportions of type I and IIB fibres and IMF content (Kim et al., 2013), although the total number of fibres and density of type IIB fibres were positively related to the IMF content (Kim et al., 2013). Henckel et al. (1997) made a similar observation in pigs, that the frequency of type IIB fibres was positively correlated to IMF. However those who have reviewed this area have concluded that there does not appear to be a clear relationship between the quantity of intramuscular adipocytes and fibre type (Lefaucheur, 2009; Hocquette In relation to the metabolic activity of type IIB fibres, these fast-twitch et al., 2010). glycolytic fibres have a higher dependence on glycolytic metabolism, therefore they have higher glycogen contents. As a result, muscles that have a high proportion of these types of fibres tend to have an increased rate of postmortem pH decline, along with a lower ultimate pH, as well as decreased water holding capacity and a paler colour (Choe et al., 2008; Larzul et al., 1997; Ryu & Kim, 2005). The fast type fibres have greater cross sectional areas (CSA) than the slow fibres, with the type IIB fibres having the greatest CSA. However muscles containing fibres with a high CSA have been reported as having a lower meat quality in both cattle (Renand et al., 2001) and pigs (Karlsson et al., 1993; Kim, et al., 2013). However the relationship between CSA per se and meat quality is not absolutely clear. Studies that have reported this relationship have often done so in animals that have variant genes or are subject to growth promoter treatments, which show a change in CSA, but this is associated with alterations in metabolic activity (Lefaucheur, 2009). For example, although callipyge lambs do have fibres with a large CSA, their muscles contain very high concentrations of calpastatin, which is thought to be a more significant factor in causing their extreme toughness (Duckett, Snowder, & Cockett, 2000).

5. Proteolytic systems and meat quality

The final tenderness of meat depends on the degree of alteration of the muscle structural and associated proteins postmortem (Hopkins & Thompson, 2002). Specific myofibrillar, myofibril cytoskeleton and costamere proteins are subjected to cleavage, with selected and restricted cleavage of the major myofibrillar proteins such as actin and myosin (Goll, Thompson, Taylor, & Christiansen, 1992; Taylor, Geesink, Thompson, Koohmaraie, & Goll, 1995a; Lametsch et al., 2003). There are several endogenous proteolytic systems present in muscle, which could participate in postmortem proteolysis, these include the cathepsin, proteasome, caspase and calpain systems (Kemp, Sensky, Bardsley, Buttery, & Parr, 2010). Generally it is considered that the cathepsin system does not play a role in meat tenderisation, as there is little association between cathepsin activity and the variation in the tenderness of meat (Whipple et al., 1990).

The proteasome is a multicatalytic protease complex involved in the regulation of a number of basic cellular pathways, by their degradation of proteins in the cytosol and nucleus (Coux, Tanaka, & Goldberg, 1996). Proteolysis by the proteasome is an ubiquitin-dependent process; poly-ubiquitinated proteins are subsequently recognised by the proteasome, which then degrades them (Taillandier et al., 2004). This process is ATP-dependent requiring a number of enzymes to ubiquitinate target proteins. In muscle there are ubiquitin system ligases which appear to be critical to the proteasomes involvement in protein degradation. These are muscle RING finger 1 (MuRF1) and muscle atrophy F-box (MAFbx) (Bodine & Baehr, 2014.). These ligases are strongly associated with muscle atrophy conditions (Glass, 2005). Although in vivo the proteasome requires ATP for target proteins to be ubiquitinated, the proteasome can carry out protoeolytic activity without requiring ATP or ubiquitin (Peters, Franke, & Kleinschmidt, 1994). Indeed proteasome activity is maintained during the postmortem conditioning period, with reported substantial activity still detectable at 7 days postmortem and at pH levels of less than 6 (Lamare, Taylor, Farout, Briand, & Briand, 2002). Taylor et al. (1995b) and Robert et al. (1999) found that the proteasome was capable of causing proteolysis of myofibril proteins including nebulin, myosin, actin and tropomyosin in bovine myofibrils. However, as emphasized by Koohmaraie and Geesink (2006), the degradation pattern of myofibrillar proteins in incubations with proteasome are not the same as that seen in postmortem muscle, although this does not appear to exclude the proteasome from making a contribution to the process of postmortem proteolysis (Houbak, Ertbjerg, & Therkildsen, 2008).

The caspase system is involved in the process of programmed cell death (apoptosis). system consists of a number of enzymes whose substrates include cytoskeletal proteins which are degraded both in apoptosis and in postmortem muscle. It has been proposed that the protease family of caspases could be active postmortem and contribute to tenderization (Sentandreu, Coulis, & Ouali, 2002; Ouali et al., 2006, Kemp et al., 2010; Kemp & Parr, 2012). Of particular interest are the effector caspases, such as caspase 3 and 7, which are activated by upstream initiator caspases, such as caspase 9, and once activated target and cleave specific substrates, resulting in cell disassembly (Fuentes-Prior & Salvesen, 2004). There have been descriptions of caspase activity as well as cleavage of indicative caspase substrates which are associated with meat tenderization, including potential effects on the calpain inhibitor, calpastatin (Kemp & Parr, 2012). However others have reported no changes in caspase activity postmortem, suggesting a lack of involvement in tenderization (Underwood, Means, & Du, 2008), whilst others have indicated associations between caspase activity and myofibril protein degradation as well as postmortem proteolysis (Huang, Huang, Xu, & Zhou, 2008; Huang, Huang, Zhou, Xu, & Xue, 2011). However whether this proteolytic system plays a major role in influencing tenderness is debatable. If it does have a role in the postmortem proteolysis which influences meat tenderness, it is likely to be contributory rather than causative.

Of all the proteolytic systems the calpain system is the one which has the strongest experimental evidence demonstrating a role in post-mortem proteolysis (Kemp et al., 2010; Koohmaraie & Geesink, 2006; Sentandreu et al., 2002). In skeletal muscle, the calpain system consists of three proteases, ubiquitously expressed isoforms Calpain-1, Calpain-2, and p94 (or Calpain-3) and the calpain-specific endogenous inhibitor, calpastatin (Goll, Thompson, Li, Wei, & Cong, 2003; Wendt, Thompson, & Goll, 2004). evidence suggests Calpain-1 has the most significant role in postmortem proteolysis and meat tenderisation. Much of this evidence has been based on observations on the association between Calpain-1 activity, as well as expression, to tenderness (Koohmaraie & Geesink, 2006). The important role of Calpain-1 in the tenderisation process was further strengthened by observations made in calpain-1 knockout mice, which indicate a direct effect of Calpain-1 in postmortem proteolysis and thereby the development of tender meat (Geesink, Kuchay, Chishti, & Koohmaraie, 2006). A consistent observation of the calpain system's involvement in tenderness is that high levels of calpastatin are associated with poor quality meat; the model being that high levels of calpastatin reduce the activity of calpain (predominantly Calpain-1) thereby reducing the proteolysis required for tender meat (Kemp et al., 2010).

Changing animal nutrition has been shown to alter calpain expression and is associated with changes in meat quality. In pigs, a moderate reduction in both protein (14.95% vs 11.08%) and digestible energy (13.81MJ vs 12.55MJ) intake has been shown to attenuate expression of Calpain-1 mRNA and this was associated with decreased Warner-Bratzler shear force (Tang et al., 2010). However, much earlier studies in lambs found that a more severe reduction in feed intake over 6 weeks (so average daily live weight gain was 14% of controls), did not alter calpain or calpastatin activity (Higgins, Lasslett, Bardsley, & Buttery, 1988). Changes in the calpain system activity have also been described associated with increased feed efficiency. Pigs selected for low residual feed intake (RFI), where their observed feed intake is lower than expected (based on body weight gain and back fat depth), have reduced protein degradation compared to those with high RFI (Cruzen et al., 2013). Relative to high RFI pigs, low RFI animals had lower proteasome activity, greater calpastatin activity and decreased Calpain-1:calpastatin activity ratio. These changes were associated with slowed postmortem muscle proteolysis, indicated by decreased Troponin T degradation. This study is consistent in showing that a decrease in the proteolytic process and subsequent reduction in meat tenderness is predominantly associated with increased inhibition of calpains. addition, this study suggests that if increased feed efficiency is achieved by a decrease in protein degradation, this could have a detrimental effect on meat quality.

Our studies on a random selection of commercially slaughtered pigs have shown that a high level of calpastatin (both activity and protein levels) in the first few hours after slaughter are associated with an increased incidence of toughness at 8 days postmortem (Sensky et al., 1998; Parr et al., 1999). The observations that Calpain-1 and calpastatin are important determinants for tender meat has lead to efforts to identify genetic markers that could be used to select for postmortem tenderisation of meat. Genetic tests for variation in both the Calpain-1 (CAPN1) and calpastatin (CAST) genes, which are associated with tenderness, have been identified for cattle (Page et al., 2004; White et al., 2005; Casas et al., 2006). In pigs, similar efforts have identified genetic markers for CAST which have been suggested to be markers of pork meat quality traits (Ciobanu et al., 2004, Nonneman et al., 2011), but as yet predictive variations in CAPN1 have not been reported.

6. Effects of BA and GH on fibre type, proteolytic systems and metabolism.

The objective of administration of the growth promoters is to ideally increase feed efficiency but at the same time increase lean tissue deposition (increase muscle mass) and decrease fat deposition, thereby repartitioning nutrients away from fat to muscle growth. Given that post-

natal muscle growth involves hypertrophy rather than hyperplasia it might be expected that stimulated growth would result in increased muscle fibre hypertrophy. Of the growth promoters BA and GH, the former has the strongest muscle growth effects. Both BA and GH have been shown to increase muscle fibre hypertrophy (Kim & Sainz, 1992), but to a more limited extent for GH (Solomon, Campbell, & Steele, 1990). In GH treated animals there appears to be no effect on fibre type distribution and this is particularly the case for pigs (Aalhus, Best, Costello, & Schaefer, 1997; Oksbjerg et al., 1995). However for animals treated with BA there are well documented effects on fibre type distributions. Early studies in lambs indicated that the BA cimaterol (10ppm, oral dose, for 5 weeks) caused an increase in type II fibres compared to type I (Beermann et al., 1987). In pigs treated over a four week period with the BA, ractopamine (20ppm, oral dose), there was a sustained induction of MyHCIIb in longissimus muscle (Gunawan, Richert, Schinckel, Grant, & Gerrard, 2007). This change in gene expression takes place early in the treatment, within one day for MyHCIIb. Likewise in pigs following seven days of treatment with ractopamine (10ppm, oral dose), in longissimus muscle there was a trend for MyHCIIb mRNA to increase and for MyHCIIa mRNA to decrease (Brown, Ryan, Daniel, Mareko, Parr, & Brameld, 2012) and this was before a significant increase in muscle weights was observed (Ryan, Daniel, Mareko, Brown, Parr, & Brameld, 2012). In the same study, pigs were also treated with GH for seven days (10mg/pig, intramuscular injection every 2 days). There was a trend for decrease in MyHCIIb gene expression in muscles with GH, but no indication of changes in muscle weights (Brown et al., 2012). Similarly in lambs, following a six day treatment with cimaterol (10ppm, oral dose), there was a decrease in MyHCIIa isoform gene expression and an increase in MyHCIIx/b, which consisted of an induction of MyHCIIb, a MyHC not normally expressed in sheep muscles (Hemmings, Daniel, Buttery, Parr, & Brameld, 2015). This switch to fast MyHC gene expression was accompanied by an increase in selective muscle weights and CSA (unpublished observations). It was interesting to note that although MyHC gene expression was altered after six days, the quantity of MyHC protein did not change (Hemmings et al., 2015). However there was a decrease in isocitrate dehydrogenase activity which suggested that there was a change to a less oxidative metabolism, before there was a change in the contractile proteins associated with a fast-glycolytic fibre type (Hemmings et al., 2015).

The net deposition of protein often results in an increase in protein synthesis, however alterations in protein degradation can also take place. Growth hormone stimulated increases in IGF-1 potentially mediates its effects through MEK/ERK and Akt–mTOR–S6K pathways.

Although IGF-1 stimulated protein synthesis can be influenced by both pathways, the effect of IGF-1 on protein degradation is mediated by the latter. The Akt–mTOR–S6K pathway also controls protein degradation in skeletal muscle by AKT phosphorylating the forkhead box protein (FOXO) transcription factors, thereby reducing the gene expression of the ubquitin ligases MAFbx and MuRF1 (Glass, 2005). These two ubiquitin ligases are important in muscle protein turnover, as they target myofibrillar proteins for degradation through the ubiquitin-proteasome system (Bodine & Baehr, 2014).

For BA, early reports on the mechanism of its action described how clenbuterol treatment for five weeks (10ppm, oral dose) significantly reduced muscle protein degradation in lambs (Bohorov et al., 1987), although subsequent studies have shown BA also stimulate protein synthesis, for example in pigs given ractopamine for four weeks (20ppm, oral dose) (Adeola et al., 1992). The characteristic effect of orally administered BA such as clenbuterol or cimaterol on protein degradation is an increase in the expression and activity of calpastatin (Higgins et al., 1988; Parr, Bardsley, Gilmour, & Buttery, 1992). This observation has been confirmed by others, particularly in ruminants (Kretchmar, Hathaway, Epley, & Dayton, 1990; Wheeler & Koohmaraie, 1992), but also in pigs (Parr, Sensky, Bardsley, & Buttery, 2001). In a more recent study, Douillard et al. (2012) described how over expression of exogenous calpastatin in mouse tibialis anterior attenuated the muscle hypertrophy and the shift to a fast muscle fibre type, stimulated by clenbuterol treatment for 21 days (subcutaneous injection, 1mg/kg body weight/day). This was in spite of AKT and associated factors being phosphorylated, an event which is normally associated with increased muscle synthesis. The authors suggested that inhibition of clenbuterol stimulated hypertrophy by calpastatin over-expression did not involve inhibition of protein synthesis. suggested that the exogenous calpastatin expression inhibited calpain's role in remodeling the muscle myofibrillar proteins. Care must be taken in accepting this interpretation, as Douillard et al. (2012) described how in control mice treated with only clenbuterol for 21 days (subcutaneous injection, 1mg/kg body weight/day) there was decreased calpastatin expression. This contradicts previous studies in rodents which have shown BA increases calpastatin in mice treated with formoterol for 28 days (intraperitoneal injection, 100microg/kg body weight/day) (Koopman et al., 2010) and rats treated with clenbuterol for 3 days (intraperitoneal injection, 3mg/kg body weight/day) (Gonçalves et al., 2011). Overall the majority of studies demonstrate treatment with BA increases muscle calpastatin. The effect of BA on normal muscle is to enhance protein synthesis and reduce calcium—dependent proteolysis, which appears to be mediated through an increase in calpastatin. However in

mice with denervated muscle, which induces atrophy, treatment with clenbuterol for 3 days (intraperitoneal injection, 3mg/kg body weight/day) stimulated protein synthesis and inhibited the proteolysis mediated by both the proteasome and lysosome, reduced expression of the ubiquitin ligases, MAFbx and MuRF1, as well as cathepsin L, without a significant effect on calcium dependent proteolysis (Gonçalves et al., 2011). This suggests that the calpain system mediated proteolysis is associated more with normal muscle protein turnover, rather than that associated with atrophy

Our recent studies have sought to examine the effects of GH or BA treatment on muscle proteolytic systems. In pigs treated with either ractopamine (20ppm, oral dose) or GH (10mg/pig, intramuscular injection every 2 days) for 27 days there was a significant effect of this BA on muscle weights and this was associated with a switch to increased expression of the MyHC isoforms associated with fast fibre types, whereas there was no effect of GH (Brameld et al., 2015). In terms of effects on meat quality, there was an indication of an increase in shear force in BA treated animals (P = 0.107). There was a trend (P < 0.1) for increased calpastatin protein (detected by western blot) in BA treated animals, but there was no effect on caspase 3/7 activity (when assessing activity these two caspases are indistinguishable) (Mareko, Ryan, Brown, Brameld, & Parr, 2013a). When proteasome activity was assessed there was a change in the activity of its subunits; these have trypsin-like, caspase-like and chymotrypsin-like activity. When these activities were assessed individually, the caspase-like activity was significantly reduced in both BA and GH treated pigs, and the chymotrypsin-like activity was increased in BA treated animals only (Mareko, Ryan, Brown, Brameld, & Parr, 2013b). Overall these data would suggest that the effect of the BA ractopamine on muscle proteolysis is not as great as some of the other BAs used in earlier studies, particularly cimaterol and clenbuterol.

In addition to examining changes in proteolytic activity in pigs treated with the GH (10mg/pig, intramuscular injection every 2 days) and ractopamine (20ppm, oral dose) for 27 days, we also examined the effect of these growth promoters on gene expression in the Longissimus muscle at days 1, 3, 7, 13 and 27 of treatment. The changes in the transcriptome were measured using a pig transcriptome microarray. Although our previous studies had indicated stimulatory effects of BAs on the gene expression of the calpain system in pigs, particularly increases in calpastatin gene expression (Parr et al., 2001), we found no consistent significant effects on gene expression of any major proteolytic system in muscle across the time course. However as might have been expected, the greatest gene expression responses were seen with ractopamine treatment. There was an increase in the expression of genes

encoding glycolysis enzymes but a decreased expression of genes associated with oxidative respiration. However the most predominant effect was a co-ordinate increase in serine synthesis pathway gene expression, with increased expression of phosphoglycerate dehydrogenase (PHGDH), phosphoserine-aminotransferase (PSAT) and phosphoserine phosphatase (PSPH). This was confirmed at the protein level, as PHGDH was significantly increased with BA (Brameld et al., 2015). As had been observed in the past, BA had a stronger muscle hypertrophic effect than GH. The effects of ractopamine on proteolytic systems do not appear to be as potent as previous described for the standard beta-2 agonists, such as cimaterol and clenbuterol. However, ractopamine does appear to be mediating its effect on muscle in pigs through biosynthetic pathways.

7. Conclusion

The consumption of meat as a protein source is predicted to increase dramatically in the next few decades. This requirement along with the increasing pressure of feed availability will demand that animal production systems are efficient. To achieve this it is very likely that the use of growth promoters will increase, as well as the possible use of genetically modified The current licensed growth promoters, which have direct effects on muscle organisms. growth as well as fat deposition, were discovered over 30 years ago. Although their effects on meat quality tend to be negative, they do not appear, at least in pigs, to have such drastic effects which cause a significant number of consumers to perceive poor meat quality. However it is likely there will be an increased incentive to develop new types of growth promoters. Fast glycolytic fibres appear to be the most receptive to growth stimuli whilst slow aerobic fibres appear to be relatively resistant to anabolic stimuli. One of the major limitations of muscle growth is that fibre number is essentially fixed at birth for most species, which means post-natal anabolic agents mediate their effects via hypertrophy. One of the consequences of hypertrophy is oxygen becomes less available, therefore glycolytic dependent metabolism becomes more favourable. Care has to be taken to ensure new types of growth promoters do not target mechanisms which result in increased CSA of fast glycolytic fibres and significant decreases in protein degradation (proteolytic systems), as there is then likely to be a significant decline in meat quality.

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References

- Aalhus, J. L., Best, D. R., Costello, F., & Schaefer, A. L. (1997). The effects of porcine somatotropin on muscle fibre morphology and meat quality of pigs of known stress susceptibility. Meat Sci,45, 283–295.
- Adeola, O., Ball, R. O., & Young, L. G. (1992). Porcine skeletal muscle myofibrillar protein synthesis is stimulated by ractopamine. Journal of Nutrition, 122, 488–495.
- Altarejos, J. Y., & Montminy, M. (2011). CREB and the CRTC co-activators: sensors for hormonal and metabolic signals. Nature Reviews. Molecular Cell Biology, *12*(3), 141–51.
- Beermann, D. H. (1994). Carcass composition of animals given partitioning agents. In H. D. Hafs & R. G. Zimbleman (Eds.), Low Fat Meats (pp. 203–222). San Diego: Academy Press, Inc.
- Beermann, D. H., Butler, W. R., Hogue, D. E., Fishell, V. K., Dalrymple, R. H., Ricks, C. A., & Scanes, C. G. (1987). Cimaterol-induced muscle hypertrophy and altered endocrine status in lambs. J Anim Sci, 65, 1514-1524.
- Bodine S.C. & Baehr, L.M. (2014) Skeletal muscle atrophy and the E3 ubiquitin ligases MuRF1 and MAFbx/atrogin-1. Am J Physiol Endocrinol Metab, 307, E469–E484.
- Bohorov, O., Buttery, P. J., Correia, J. H. R. D., & Soar, J. B. (1987). The effect of the B-2-adrenergic agonist clenbuterol or implantation with oestradiol plus trenbolone acetate on protein metabolism in wether lambs. British Journal of Nutrition, 57, 99–107.
- Boisclair, Y. R., Rhoads, R. P., Ueki, I., Wang, J., & Ooi, G. T. (2001). The acid-labile subunit (ALS) of the 150 kDa IGF-binding protein complex: An important but forgotten component of the circulating IGF system. Journal of Endocrinology, 170, 63-70.
- Brameld, J. M., Atkinson, J. L., Saunders, J. C., Pell, J. M., Buttery, P. J., & Gilmour, R. S. (1996). Effects of growth hormone administration and dietary protein intake on insulinlike growth factor I and growth hormone receptor mRNA expression in porcine liver, skeletal muscle, and adipose tissue. J Anim Sci, 74, 1832-1841.
- Brameld, J., Ryan, K., Williams, H., Harris, D., Brown, D., Emes, R., ... Parr, T. (2015). Transcriptomic and metabolomic assessment of growth promoter effects on porcine muscle growth. J Anim Sci, 93, Suppl. s3, 223.

- Brooke, M. H., & Kaiser, K. K. (1970). Muscle fiber types How many and what kind. Archives of Neurology, 23, 369–379.
- Brown, D., Ryan, K., Daniel, Z., Mareko, M. H. D., Parr, T., & Brameld, J. (2012). Beta-adrenergic agonist and growth hormone have differential effects on muscle fibre composition in growing gilts. Proceedings of the British Society of Animal Science Vol 3(1): pp66.
- Bush, J. A., Burrin, D. G., Suryawan, A., O'Connor, P. M., Nguyen, H. V., Reeds, P. J., . . . Davis, T. A. (2003). Somatotropin-induced protein anabolism in hindquarters and portal-drained viscera of growing pigs. Am J Physiol Endocrinol Metab, 284(2), E302-312.
- Casas, E., White, S.N., Wheeler, T.L., Shackelford, S.D., Koohmaraie, M., Riley, D.G., ... Smith, T.P. (2006). Effects of calpastatin and u-calpain markers in beef cattle on tenderness traits. J Anim Sci, 84, 520-525.
- Choe, J. H., Choi, Y. M., Lee, S. H., Shin, H. G., Ryu, Y. C., Hong, K. C., & Kim, B. C. (2008). The relation between glycogen, lactate content and muscle fiber type composition, and their influence on postmortem glycolytic rate and pork quality. Meat Sci, 80(2), 355-362.
- Ciobanu, D.C., Bastiaansen, J.W.M., Lonergan, S.M., Thomsen, H., Dekkers, J.C.M., Plastow, G.S., & Rothschild, M.F. (2004). New alleles in calpastatin gene are associated with meat quality traits in pigs. J Anim Sci, 82, 2829-2839.
- Clemmons, D. R. (2009). Role of IGF-I in skeletal muscle mass maintenance. Trends in Endocrinology and Metabolism, 20: 349-356.
- Coux, O., Tanaka, K., & Goldberg, A. L. (1996). Structure and functions of the 20S and 26S proteasome. Annual Review of Biochemistry, 65, 801–847.
- Cruzen, S. M., Harris, A. J., Hollinger, K., Punt, R. M., Grubbs, J. K., Selsby, J. T., . . . Huff-Lonergan, E. (2013). Evidence of decreased muscle protein turnover in gilts selected for low residual feed intake. J Anim Sci, 91, 4007-4016.
- Daughaday, W. H. (2000). Growth hormone axis overview somatomedin hypothesis. Pediatr. Nephrol. 14:537-540.
- Denley, A., Cosgrove, L. J., Booker, G. W., Wallace, J. C., & Forbes, B. E. (2005). Molecular interactions of the IGF system. Cytokine Growth Factor Rev. 16:421-439.
- Etherton, T. D., & Bauman, D. E. (1998). Biology of somatotropin in growth and lactation of domestic animals. Physiology Reviews, 78, 745–761.
- Douillard, A., Galbes, O., Begue, G., Rossano, B., Levin, J., Vernus, B., . . . Py, G. (2012). Calpastatin overexpression in the skeletal muscle of mice prevents clenbuterol-

- induced muscle hypertrophy and phenotypic shift. Clin Exp Pharmacol Physiol, 39, 364-372.
- Duckett, S. K., Snowder, G. D., & Cockett, N. E. (2000). Effect of the callipyge gene on muscle growth, calpastatin activity, and tenderness of three muscles across the growth curve. J Anim Sci, 78, 2836–2841.
- Dunshea, F. R., Cox, M. L., Borg, M. R., Sillence, M. N., & Harris, D. R. (2002). Porcine somatotropin (pST) administered using a commercial delivery system improves growth performance of rapidly-growing, group-housed finisher pigs. Australian Journal of Agricultural Research, 53, 287–293.
- Dunshea, R. R., D'Souza, D. N., Pethic, D. W., Harper, G. S., & Warner, R. D. (2005). Effects of dietary factors and other metabolic modifiers on quality and nutritional value of meat. Meat Sci, 71, 8–38.
- Etherton, T. D., & Bauman, D. E. (1998). Biology of somatotropin in growth and lactation of domestic animals. Physiology Reviews, 78,745–761.
- Etherton, T. D., Wiggins, J. P., Evock, C. M., Chung, C. S., Rebhun, J. F., Walton, P. E., & Steele, N. C. (1987). Stimulation of pig growth performance by porcine growth hormone: determination of the dose-response relationship. J Anim Sci, 64, 433-443.
- FAO (2009). How to Feed the World in 2050. High-Level Expert Forum, Rome 12-13th
 October 2009.
 http://www.fao.org/fileadmin/templates/wsfs/docs/expert_paper/How_to_Feed_the_Worl

d_in_2050.pdf

- Foresight. (2011). The Future of Food and Farming: Challenges and choices for global sustainability. Final Project Report. The Government Office for Science, London. https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/288329/11 546-future-of-food-and-farming-report.pdf
- Geesink, G.H., Kuchay, S., Chishti, A.H., & Koohmaraie, M. (2006). Micro-calpain is essential for postmortem proteolysis of muscle proteins. J Anim Sci, 84, 2834-2840.
- Glass, D. J. (2005). Skeletal muscle hypertrophy and atrophy signaling pathways. The International Journal of Biochemistry & Cell Biology, *37*(10), 1974–84.
- Godfray, H. C., Beddington, J. R., Crute, I. R., Haddad, L., Lawrence, D., Muir, J. F., . . . Toulmin, C. (2010). Food security: the challenge of feeding 9 billion people. Science, 327(5967), 812-818.
- Goll, D.E., Thompson, V.F., Li, H.Q., Wei, W., & Cong, J.Y. (2003). The calpain system. Physiological Reviews, 83, 731-801.

- Goll, D.E., Thompson, V.F., Taylor, R.G., & Christiansen, J.A. (1992). Role of the calpain system in muscle growth. Biochimie, 74, 225-237.
- Goncalves, D. A., Silveira, W. A., Lira, E. C., Graca, F. A., Paula-Gomes, S., Zanon, N. M., . . . Navegantes, L. C. (2012). Clenbuterol suppresses proteasomal and lysosomal proteolysis and atrophy-related genes in denervated rat soleus muscles independently of Akt. Am J Physiol Endocrinol Metab, 302, E123-133.
- Gunawan, A. M., Richert, B. T., Schinckel, A. P., Grant, A. L., & Gerrard, D. E. (2007).

 Ractopamine induces differential gene expression in porcine skeletal muscles. J Anim Sci, 85, 2115–2124.
- Hemmings, K.M., Daniel, Z.C., Buttery, P.J., Parr, T., & Brameld, J.M. (2015) Differential effects of short-term β agonist and growth hormone treatments on expression of myosin heavy chain IIB and associated metabolic genes in sheep muscle. Animal, 9, 285-94.
- Henckel, P., Oksbjerg, N., Erlandsen, E., Barton-Gade, P., & Bejerholm, C. (1997). Histoand biochemical characteristics of the longissimus dorsi muscle in pigs and their relationships to performance and meat quality. Meat Sci, 47, 311–321.
- Higgins, J. A., Lasslett, Y. V., Bardsley, R. G., & Buttery, P. J. (1988). The relation between dietary restriction or clenbuterol (a selective beta 2 agonist) treatment on muscle growth and calpain proteinase (EC 3.4.22.17) and calpastatin activities in lambs. Br J Nutr, 60, 645-652.
- Hocquette, J. F., Gondret, F., Baeza, E., Medale, F., Jurie, C., & Pethick, D. W. (2010). Intramuscular fat content in meat-producing animals: Development genetic and nutritional control and identification of putative markers. Animal, 4, 303–319.
- Houbak, M.,B., Ertbjerg, P., & Therkildsen, M. (2008). In vitro study to evaluate the degradation of bovine muscle proteins post-mortem by proteasome and micro-calpain.
 Meat Sci, 79, 77–85. Huang, M., Huang, F., Xu, X., & Zhou, G. (2008). Influence of caspase3 selective inhibitor on proteolysis of chicken skeletal muscle proteins during post mortem aging. Food Chemistry, 115, 181–186.
- Huang, F., Huang, M., Zhou, G., Xu, X., & Xue, M. (2011). In vitro proteolysis of
 myofibrillar proteins from beef skeletal muscle by caspase-3 and caspase-6. Journal of
 Agricultural and Food Chemistry, 59, 9658–9663.
- Karlsson, A., Enfalt, A. C., Essen-Gustavsson, B., Lundstrom, K., Rydhmer, L., & Stern, S. (1993). Muscle histochemical and biochemical properties in relation to meat qual- ity during selection for increased lean tissue growth rate in pigs. J Anim Sci, 71, 930–938.

- Kemp, C. M., & Parr, T. (2012). Advances in apoptotic mediated proteolysis in meat tenderisation. Meat Sci, 92, 252–9.
- Kemp, C. M., Sensky, P. L., Bardsley, R. G., Buttery, P. J., & Parr, T. (2010). Tenderness -an enzymatic view. Meat Sci, 84, 248–56.
- Kim, G. D., Jeong, J. Y., Jung, E. Y., Yang, H. S., Lim, H. T., & Joo, S. T. (2013). The influence of fiber size distribution of type IIB on carcass traits and meat quality in pigs. Meat Sci, 94, 267–273.
- Kim Y.S. & Sainz R.D. (1992) Beta-adrenergic agonists and hypertrophy of skeletal muscles. Life Science, 50, 397-407.
- Koohmaraie, M., & Geesink, G.H. (2006). Contribution of postmortem muscle biochemistry to the delivery of consistent meat quality with particular focus on the calpain system.

 Meat Sci, 74, 34-43.
- Koopman, R., Gehrig, S. M., Leger, B., Trieu, J., Walrand, S., Murphy, K. T., & Lynch, G. S. (2010). Cellular mechanisms underlying temporal changes in skeletal muscle protein synthesis and breakdown during chronic beta-adrenoceptor stimulation in mice. J Physiol, 588, 4811-4823.
- Kretchmar, D. H., Hathaway, M. R., Epley, R. J., & Dayton, W. R. (1990). Alterations in postmortem degradation of myofibrillar proteins in muscle of lambs fed a b-adrenergic agonist. J Anim Sci, 68, 1760–1772.
- Krick, B. J., Roneker, K. R., Boyd, R. D., Beermann, D. H., David, P. J., & Meisinger, D. J. (1992). Influence of genotype and sex on the response of growing pigs to recombinant porcine somatotropin. J Anim Sci, 70, 3024–3034.
- Lamare, M., Taylor, R.G., Farout, L., Briand, Y., & Briand, M. (2002). Changes in proteasome activity during postmortem aging of bovine muscle. Meat Sci, 61, 199-204.
- Lametsch, R., Karlsson, A., Rosenvold, K., Andersen, H.J., Roepstorff, P., & Bendixen, E. (2003). Postmortem proteome changes of porcine muscle related to tenderness. Journal of Agricultural and Food Chemistry, 51, 6992-6997.
- Larzul, C., Lefaucheur, L., Ecolan, P., Gogue, J., Talmant, A., Sellier, P., . . . Monin, G. (1997). Phenotypic and genetic parameters for longissimus muscle fiber characteristics in relation to growth, carcass, and meat quality traits in large white pigs. J Anim Sci, 75, 3126-3137.
- Lefaucheur, L. (2010). A second look into fibre typing Relation to meat quality. Meat Sci, 84, 257–270.

- Liang, W., & Mills, S. E. (2002). Quantitative analysis of β-adrenergic receptor subtypes in pig tissues. J Anim Sci, 80, 963–970,
- Mareko, M. H. D., Ryan, K., Brown, D., Brameld, J., & Parr, T. (2013a). Effects of 27 days of treatment with growth promoting agents on adult pig *longissimus* muscle protease systems and post-mortem shear force after conditioning. Proceedings of the British Society of Animal Science Vol 4(1): pp151.
- Mareko, M. H. D., Ryan, K., Brown, D., Brameld, J., & Parr, T. (2013b). Effects on pork quality of gilts treated with Ractopamine or growth hormone for 7 days and 27 days. Proceedings of the International Conference on Meat Science and Technology (59th ICoMST 2013). Izmir, Turkey, 18 23rd August, 2013, Abstracts S4 7.
- Meersman, H.J. (1998). Overview of the Effects of beta-Adrenergic Receptor Agonists on Animal Growth Including Mechanisms of Action. J Anim Sci, 76, 160-172.
- Mills, S.E., Kissel, J., Bidwell, C.A., & Smith, D.J. 2003. Stereoselectivity of porcine beta-adrenergic receptors for ractopamine stereoisomers. J Anim Sci, 81, 122–129.
- McNeel, R.L., & Mersmann, H.J. (1995). Beta-adrenergic receptor subtype transcripts in porcine adipose tissue. J Anim Sci, 73, 1962-71.
- Nonneman, D., Lindholm-Perry, A. K., Shackelford, S. D., King, D. A., Wheeler, T. L., Rohrer, G. A., . . . Moeller, S. J. (2011). Predictive markers in calpastatin for tenderness in commercial pig populations. J Anim Sci, 89, 2663-2672.
- Oksbjerg, N., Petersen, J. S., Sorensen, M. T., Henckel, P., Agergaard, N., Bejerholm, C., & Erlandsen, E. (1995). The influence of porcine growth hormone on muscle fibre characteristics, metabolic potential and meat quality. Meat Sci, 39, 375-385.
- Ouali, A., Herrera-Mendez, C. H., Coulis, G., Becila, S., Boudjellal, A., Aubry, L., & Sentandreu, M. A. (2006). Revisiting the conversion of muscle into meat and the underlying mechanisms. Meat Sci, 74, 44-58.
- Page, B. T., Casas, E., Quaas, R. L., Thallman, R. M., Wheeler, T. L., Shackelford, S. D., . . . Smith, T. P. (2004). Association of markers in the bovine CAPN1 gene with meat tenderness in large crossbred populations that sample influential industry sires. J Anim Sci, 82, 3474-3481.
- Parr, T., Bardsley, R.G., Gilmour, R.S., & Buttery, P.J. (1992). Changes in Calpain and Calpastatin mRNA Induced by Beta-adrenergic Stimulation of Bovine Skeletal Muscle. European Journal of Biochemistry, 208, 333-339.

- Parr, T., Sensky, P. L., Bardsley, R. G., & Buttery, P. J. (2001) Calpastatin Expression in Cardiac and Skeletal Muscle and Partial Gene Structure. Archives of Biochemistry and Biophysics 395, 1-13
- Parr, T., Sensky, P. L., Scothern, G., Bardsley, R. G., Buttery, P. J., Wood, J. D. & Warkup,C. C. (1999). Immunochemical study of the calpain system in porcine longissimusmuscle with high and low shear force values. J Anim Sci, 77 (suppl 1): 164.
- Patience, J.F., Rossoni-Serao, M.C., & Gutierrez, N.A. (2015). A review of feed efficiency in swine: biology and application. Journal of Animal Science and Biotechnology, 6, 33..
- Peters, J. M., Franke, W. W., & Kleinschmidt, J. A. (1994). Distinct 19S and 20S subcomplexes of the 26S proteasome and their distribution in the nuleus and the cytoplasm. The Journal of Biological Chemistry, 269, 7709–771
- Renand, G., Picard, B., Touraille, C., Berge, P., & Lepetit, J. (2001). Relationships between muscle characteristics and meat quality traits of young Charolais bulls. Meat Sci, 59, 49–60.
- Ryan, K., Daniel, Z., Mareko, M. H. D., Brown, D., Parr, T., & Brameld, J. (2012). Effects of

 1 week treatment with beta-adrenergic agonist or growth hormone on the growth and feed
 conversion efficiency of growing gilts. Proceedings of the British Society of Animal
 Science Vol 3(1): pp54
- Ryu, Y. C., & Kim, B. C. (2005). The relationship between muscle fiber characteristics, postmortem metabolic rate, and meat quality of pig longissimus dorsi muscle. Meat Sci, 71, 351–357.
- Schiaffino, S., & Reggiani, C. (1996). Molecular diversity of myofibrillar proteins: Gene regulation and functional significance. Physiological Reviews, 76, 371–423.
- Sensky, P.L., Parr, T., Scothern, G.P., Perry, A., Bardsley, R.G., Buttery, P.J., ... Warkup, C. (1998). Differences in the calpain enzyme system in tough and tender samples of porcine longissimus dorsi. Proceedings of the British Society of Animal Science, (1998), 14.
- Sentandreu, M.A., Coulis, G., & Ouali, A. (2002). Role of muscle endopeptidases and their inhibitors in meat tenderness. Trends in Food Science & Technology, 13, 400-421.
- Seppelt, R., Manceur, A.M., Liu, J., Fenichel, E. P., & Klotz, S. (2014). Synchronized peakrate years of global resources use. Ecology and Society 19, 50. Siegel, P.B. (2014). Evolution of the modern broiler and feed efficiency. Annual Review of Animal Biosciences, 2, 375-385.
- Siegel, P.B. (2014) Evolution of the modern broiler and feed efficiency. Annual Review of Animal Biosciences, 2, 375-385.

- Solomon, M. B., Campbell, R. G., & Steele, N. C. (1990). Effect of sex and exogenous porcine somatotropin on longissimus muscle fiber characteristics of growing pigs. J Anim Sci, 68, 1176–1181.
- Taillandier, D., Combaret, L., Pouch, M..N., Samuels, S.E., Bechet, D., & Attaix, D. (2004). The role of ubiquitin-proteasome-dependent proteolysis in the remodelling of skeletal muscle. Proceedings of the Nutrition Society, 63, 357-361.
- Tang, R., Yu, B., Zhang, K., Guo, X., Tian, G., Huang, Z., . . . Chen, D. (2010). Effects of nutritional level on pork quality and gene expression of micro-calpain and calpastatin in muscle of finishing pigs. Meat Sci, 85, 768-771.
- Taylor, R.G., Geesink, G.H., Thompson, V.F., Koohmaraie, M., & Goll, D.E. (1995a). Is Z-disk degradation responsible for postmortem tenderization? J Anim Sci, 73, 1351-1367.
- Taylor, R.G., Tassy, C., Briand, M., Robert, N., Briand, Y., & Ouali A. (1995b). ProteolyticActivity of Proteasome on Myofibrillar Structures. Molecular Biology Reports, 21, 71-73.
- Tomas, F. M., Campbell, R. G., King, R. H., Johnson, R. J., Chandler, C. S., & Taverner, M. R. (1992). Growth hormone increases whole-body protein turnover in growing pigs. J Anim Sci, 70, 3138–3143.
- Underwood, K. R., Means, W. J., & Du, M. (2008). Caspase 3 is not likely involved in the postmortem tenderization of beef muscle. J Anim Sci, 86, 960–966.
- Velloso, C.P. (2008). Regulation of muscle mass by growth hormone and IGF-1. British Journal of Pharmacology, 154, 557-568.
- Vann, R. C., Nguyen, H. V., Reeds, P. J., Burrin, D. G., Fiorotto, M. L., Steele, N. C., . . . Davis, T. A. (2000). Somatotropin increases protein balance by lowering body protein degradation in fed, growing pigs. Am J Physiol Endocrinol Metab, 278, E477-483.
- Wang, Y., Bikle, B.D., & Chang, W. (2013). Autocrine and paracrine actions of IGf-1 signaling in skeletal development. Bone Research, 3. 249-259.
- Wendt, A., Thompson, V.F., & Goll, D.E. (2004). Interaction of calpastatin with calpain: a review. Biological Chemistry, 385, 465-472.
- Wheeler, T. L., & Koohmaraie, M. (1992). Effects of the beta adrenergic agonist L644,969 on muscle protein turnover, endogenous proteinase activities, and meat tenderness in steers.
 J Anim Sci, 70, 3035–3043
- Whipple, G., Koohmaraie, M., Dikeman, M.E., Crouse, J.D., Hunt, M.C., & Klemm, R.D. (1990). Evaluation of attributes that affect longissimus muscle tenderness in Bos taurus and Bos indicus cattle. J Anim Sci, 68, 2716-2728.

White, S. N., Casas, E., Wheeler, T. L., Shackelford, S. D., Koohmaraie, M., Riley, D. G., . . . Smith, T. P. (2005). A new single nucleotide polymorphism in CAPN1 extends the current tenderness marker test to include cattle of Bos indicus, Bos taurus, and crossbred descent. J Anim Sci, 83, 2001-2008.

The impact of growth promoters on muscle growth and the potential consequences for meat quality.

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Highlights

- The impact of the growth promoters on muscle growth
- Impacts of fibre type and proteolysis on meat quality
- Effects of growth promoters on fibre type, proteolysis and metabolism